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Application of N-PLS calibration to the simultaneous determination of Cu^{2+} , Cd^{2+} and Pb^{2+} using peptide modified electrochemical sensors

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The simultaneous determination of Cu^{2+} , Cd^{2+} and Pb^{2+} is demonstrated at four modified gold electrodes using N-PLS calibration. Three of the electrodes were modified with the peptides Gly-Gly-His, γ -Glu-Cys Gly and human angiotensin I which were covalently attached to thioctic acid self-assembled monolayers and the fourth electrode was modified with thioctic acid only. Voltammetry at the modified electrodes in the presence of the three metal ions revealed one peak due to the reduction of copper and another due to the overlapping peaks of cadmium and lead which made quantification using conventional methods difficult. N-PLS was used to calibrate and predict trace concentrations (100 nM to 10 μM) of mixtures of Cu^{2+} , Cd^{2+} and Pb^{2+} .

Introduction

Classical methods such as atomic absorption spectroscopy (AAS) and inductively coupled plasma-mass spectrometry (ICP-MS) are highly accurate and reliable for the analysis of metal ions. However, the high cost of instrumentation and complexity of operation have challenged researchers to develop new techniques to better detect and quantify metals in a variety of matrices. Stripping analysis is one of the most sensitive electroanalytical techniques for monitoring metal ions.¹ Its high sensitivity is attributed to the preconcentration step in which metals are accumulated at the working electrode. Potential scanning in a positive direction (anodic stripping voltammetry) or by applying a positive oxidative current (potentiometric stripping analysis) then allows the metal to be stripped and quantified. Work by Wang's group on screen-printed electrodes for stripping analysis² has shown the utility of the method for field applications^{3,4}. The use of square wave anodic stripping voltammetry has been demonstrated by Palchetti *et al.*⁵ in which screen-printed graphite electrodes modified with a cellulose-derivative mercury

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coating were used for the determination of lead, cadmium and copper. The stripping method provides useful information on the total metal content with detection limits at the ppb level. Perhaps more useful for biological and environmental measurements are biosensors since the biological recognition component (located in close proximity to a transducer which can be either electrochemical, optical or gravimetric) could provide information on how metal ions interact with organisms and hence bioavailable content.⁶ Biosensors can be either highly specific towards a particular analyte⁷ or display a broad range of selectivities.^{8,9} For example, Li and Lu⁷ have exploited catalytically active DNA as a biosensor for Pb^{2+} with over 80-fold selectivity over other metal ions. Hybridisation of the fluorophore labelled DNA with the complementary enzyme strand resulted in quenching of the fluorescence. In the presence of Pb^{2+} , quenching was eliminated and this intensified the fluorescence output. In a less selective approach, Csöregi and co-workers^{8,9} developed a conductivity sensor based upon a change in conformation of a metallothionein protein upon metal ion binding. The protein, GST-SmtA, was over-expressed in *Escherichia coli* and covalently attached to thiocetic acid on a gold substrate for the detection of heavy metal ions. Upon recognition of the metal ion, the protein folded to form a more compact structure and the resultant capacitance of the electrode surface increased. GST-SmtA electrodes displayed a broad selectivity for Cu^{2+} , Cd^{2+} , Hg^{2+} and Zn^{2+} with detection limits at the femtomolar range. The drawback with these two extreme types of biosensors is that in the highly specific case, detection is limited to one analyte, whereas in the other case, low selectivity does not allow the determination of complex mixtures. Recently, sensor arrays, where each particular element in the array has some selectivity towards a range of analytes, have been introduced to overcome this problem.^{10,11} The coating on each particular element of the array is chosen to provide different patterns of responses. These patterns can be deconvoluted using a chemometrics approach to obtain information about the individual components.¹²

Multivariate calibration is a useful technique for building a model of the relationship between independent and dependent variables. The calibration is posed in inverse mode with Y as the independent variable (e.g. concentration) and X as the dependent variable (e.g. voltammetric currents). The developed model is then used on new X data to predict unknown Y value(s).

Partial least squares (PLS) regression¹³⁻²⁰ is one of the more popular multivariate calibration procedures owing to its good prediction ability and ease of use. It follows from the fact that PLS searches for a compromise between better fitting (less error in X) and better prediction (less error in Y). In other words, the PLS model fitness is somewhat sacrificed in favour of its prediction ability. The PLS model can be viewed as consisting of “outer relations” of individual X and Y blocks and an “inner relation” linking both X and Y .^{13,21} This can be represented graphically as shown in Fig. 1. The goal is to

minimise the error matrix F while maintaining the correlation between X and Y through the inner relation $U = BT$.

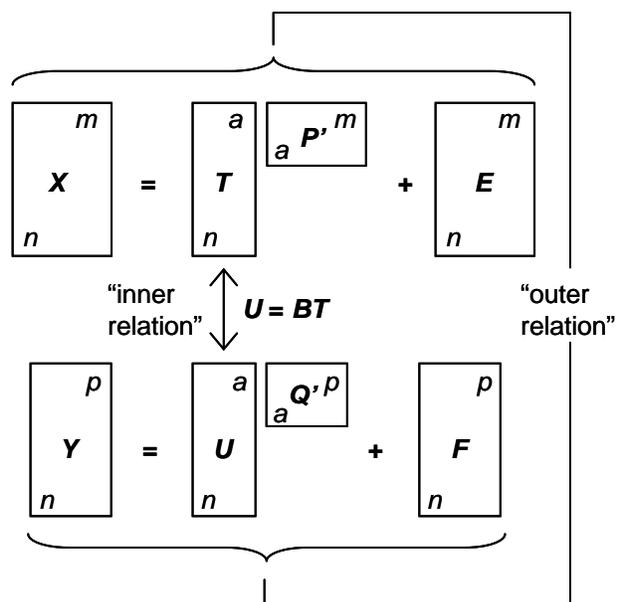


Figure 1 PLS model consisting of “outer relations” (X and Y blocks) and an “inner relation” linking both blocks.

The matrix X is decomposed into a scores matrix, T , loadings matrix, P' , and an error matrix, E . The matrix, Y , is decomposed into a scores matrix, U , loadings matrix, Q' , and an error matrix, F .¹³ The extracted factors, T , are used to predict the Y scores, U , and these predictions are then used to construct predictions for the responses.

N-PLS regression is the extension of ordinary PLS (i.e. two way PLS) to three or more (N) kinds of variables or objects.²² Two way data (e.g. voltammetric currents) can typically be represented in a data matrix with columns as variables (e.g. potential) and rows as objects (e.g. samples). By introduction of an extra mode (in the present example, different electrodes) we now have three-way data, which can be represented as a three way array (i.e. a box).

The theory and mathematical details of different algorithms for fitting and calibrating multi-way data sets has been given elsewhere¹⁹ and reiterating the literature is not aimed in this manuscript; however the algorithm chosen here is known as N-PLS which has been developed by Bro.¹⁹ It is a sequential method that decomposes the data array X similar to Parallel Factor Analysis (PARAFAC)²³ and then predicts Y . The reason to utilise a multi-linear (three-way in this case) calibration method in this study was the multi-way (three-way in this case: electrode \times potential \times solution) structure of voltammetric current data and the need to predict the concentration of three metal species.

In our work, we elect to use peptides as biorecognition molecules. Peptides are most compatible with the construction of arrays since the affinity for a particular metal ion can be easily tuned by changing the peptide sequence.^{24, 25} With twenty naturally occurring amino acids, there are many possible peptide sequences to form complexes with metals. Our research group has previously developed a simple, generic strategy towards the fabrication of peptide modified electrochemical sensors.^{26, 27} This approach involves the attachment of the amine end of a peptide to a carboxyl terminated self-assembled monolayer (SAM) on a gold electrode. The advantage of using electrochemistry is that the different redox potentials of metals can be exploited for enhanced selectivity. In earlier work, the copper binding tripeptide, Gly-Gly-His, was attached to a SAM of 3-mercaptopropionic acid (MPA) with low detection limits for Cu^{2+} .²⁷⁻²⁹ Cu^{2+} was bound to Gly-Gly-His upon accumulation in copper solution at open circuit potential for 10 minutes and was quantified by reduction of complexed Cu^{2+} to form underpotential deposited (UPD) copper by Osteryoung square wave voltammetry (OSWV).²⁹ The success of this strategy has also led to the development of sensors for Cd^{2+} using γ -Glu-Cys-Gly (glutathione)³⁰ and His-Ser-Gln-Lys-Val-Phe³¹ coupled to MPA SAMs on gold based on the UPD/stripping principle. For Pb^{2+} detection, a thioctic acid (TA) SAM was used to link human angiotensin I (Asp-Arg-Val-Tyr-Ile-His-Pro-Phe-His-Leu). Thioctic acid was necessary for stable $\text{Pb}^{2+}/\text{Pb}^0$ electrochemistry although the sensitivity was lower than that at an MPA-angiotensin I modified electrode.³² Although the nature of the peptide can result in high selectivity towards a given ion (e.g. Gly-Gly-His and Cu^{2+}), a peptide can often bind one or two other ions to a measurable extent. As a result, multiple peaks in the voltammogram were observed in the presence of electroactive interfering ions and it was difficult to quantify the amount of the analyte accurately unless there was prior knowledge of the amount of interferent present.

In this paper, we describe a peptide based electrochemical sensor array for the simultaneous determination of Cu^{2+} , Cd^{2+} and Pb^{2+} . The sensor array consists of the peptides identified in the earlier work for Cu^{2+} , Cd^{2+} and Pb^{2+} involving Gly-Gly-His, glutathione and angiotensin I, respectively. The design uses thioctic acid SAMs to link each peptide to the gold surface. Additionally, a fourth element in the array, modified with thioctic acid only was used. For calibration, the concentrations of Cu^{2+} , Cd^{2+} and Pb^{2+} were chosen to encompass the dynamic range of the sensors using an experimental design. The resulting voltammograms were calibrated using N-PLS and the ability to predict test samples was investigated.

Experimental section

Materials

Gly-Gly-His, glutathione (reduced form, >98% purity), human angiotensin I, *D,L*-6,8-thioctic acid (TA) and *N*-hydroxysuccinimide (NHS) were purchased from Sigma

(Sydney, Australia). 1-ethyl-3-(3-dimethylaminopropyl) carbodiimide hydrochloride (EDC), 2-(N-morpholino)-ethanesulfonic acid (MES), and perchloric acid were from Aldrich (Sydney, Australia). Sodium hydroxide, sodium chloride, sulfuric acid, nitric acid, ammonium acetate, ethanol, copper(II) sulfate and lead(II) nitrate were obtained from Ajax (Sydney, Australia). Cadmium(II) nitrate was purchased from Fluka (Sydney, Australia).

All solutions were prepared with Milli-Q water (18 M Ω cm, Millipore, Sydney). Buffer solutions used in this work were 50 mM ammonium acetate (pH 7.0) and 0.1 M MES (pH 6.8). The pH was adjusted with either NaOH or HNO₃ solutions. Stock metal solutions (0.1 M) were prepared in Milli-Q water and dilute metal solutions by serial dilution in ammonium acetate. All glassware was rinsed with 6 M HNO₃ followed by Milli-Q water before use to avoid metal contamination.

Preparation of modified electrodes

Gold electrodes were prepared by sealing polycrystalline gold wire (>99.99% gold, Aldrich) in 4 mm diameter glass tubes with EPON Resin 825 and EPI-CURE curing agent 3271 from Shell (Sydney, Australia).³³ The cut end of the wire was polished with 1.0 μm alumina, followed by 0.3 and 0.05 μm alumina slurry on microcloth pads (Buehler, Lake Bluff, IL). After removal of trace alumina from the surface, by rinsing with Milli-Q water and brief cleaning in an ultrasonic bath, the electrodes were further cleaned by cycling between -0.3 V and +1.5 V in 50 mM H₂SO₄ at a scan rate of 0.15 V s⁻¹ until a reproducible scan was obtained. The electrochemical area of the electrode was determined from the reduction of gold oxide by the method of Hoogvliet *et al.* using a conversion factor of 482 $\mu\text{C cm}^{-2}$.³⁴

Gold electrodes were modified with TA-Gly-Gly-His, TA-glutathione, TA-angiotensin I and TA (Fig. 2). Modification of the electrode with TA was performed by incubating a gold electrode overnight in 10 mM TA in 75% ethanol, 25% water. This was followed by rinsing with absolute ethanol.

**Scheme 1**

Steps in the fabrication of peptide modified sensors. A clean gold electrode is modified with a self-assembled monolayer of TA followed by activation of the carboxyl groups with EDC and NHS. Coupling of the *N*-terminus of the peptide results in the formation of the TA-peptide modified sensor.

Modification of the electrodes with peptides was performed as shown in Scheme 1. A TA SAM was formed on a gold electrode, followed by rinsing with ethanol and activating the carboxyl terminus by immersing in a stirred solution of 20 mM EDC and 4

mM NHS in 100 mM MES (pH 6.8) for 1 hour. After thorough rinsing with 25 mM MES buffer, the modified electrode was reacted overnight with the peptide in MES buffer to form the TA-peptide modified electrode. For modification with the peptides Gly-Gly-His and glutathione a concentration of 25 mg mL^{-1} was used, whereas for modification with angiotensin I a concentration of 10 mg mL^{-1} was used.

Figure 2: Gold electrodes modified with TA-Gly-Gly-His, TA-glutathione, TA-angiotensin I and TA.

Measurement procedure

All measurements were made in a water-jacketed cell at 25°C after equilibration for at least one hour. Copper(II), cadmium(II) and lead(II) ions were accumulated at the TA-Gly-Gly-His, TA-glutathione, TA-angiotensin I and TA modified electrodes at open circuit potential by immersing all four electrodes into 10 mL of a stirred aqueous solution of a mixture of the metal ions in 50 mM ammonium acetate (pH 7.0) for 10 min. The electrodes were then removed, rinsed with metal-free ammonium acetate and transferred to a cell with electrolyte of 50 mM ammonium acetate (pH 7.0) and 50 mM NaCl for electrochemical measurements by OSWV. After the measurement, bound metal ions were eliminated from the electrodes at $+0.5 \text{ V}$ for 30 s in 0.1 M HClO_4 .

Electrochemical measurements

All electrochemical measurements were performed with an Autolab PGSTAT 12 potentiostat (Eco Chemie, Netherlands). Multiplexing of the working electrodes was accomplished using a 16 channel switch system of which 4 were used. The four working electrodes were plugged into the inputs of the multiplexer and the output of the multiplexer was connected to the working electrode lead of the Autolab. The instrument was controlled and automated with a program written in GPES software. OSWV and time base experiments were carried out at the working electrodes with a platinum flag

auxiliary electrode and a Ag | AgCl | 3.0 M NaCl reference electrode. OSW voltammograms for the reduction process were acquired at each working electrode sequentially (TA-Gly-Gly-His, followed by TA-glutathione, TA-angiotensin I and TA) between +0.40 and -0.45 V at a pulse amplitude of 0.025 V, a step of 0.004 V and frequency of 25 Hz. The duty cycle of the multiplexer was 42 s (2 s quiet time then 8.5 s for each OSWV).

Calibration and test sets

A 16 sample set was constructed for calibration with N-PLS modelling. The analyte concentrations of Cu^{2+} , Cd^{2+} and Pb^{2+} corresponded to a central composite design: a full factorial design at two levels ($2^3 = 8$ samples), a star design ($2 \times 3 = 6$ samples) and a centre point (2 replicates). Each sample contained one of five different concentration levels (relative concentrations = -1.68, -1, 0, +1 and +1.68) of each metal ion as shown in Table 1. The order of experiments was randomised to confound uncontrolled factors. The concentration of the analytes were in the range of 10 nM to 1 μM for Cu^{2+} , 0.5 μM to 10 μM for Cd^{2+} and 50 nM to 1 μM for Pb^{2+} and were chosen to cover the dynamic range of the sensors.

Three test samples were prepared at analyte concentrations different from those chosen for calibration but within the design parameters (see Table 2).

Table 1 Coded levels and corresponding concentrations of Cu²⁺, Cd²⁺ and Pb²⁺ used in a central composite design.

Calibration solution	Level of Cu ²⁺ (<i>c</i> / μmol L ⁻¹)	Level of Cd ²⁺ (<i>c</i> / μmol L ⁻¹)	Level of Pb ²⁺ (<i>c</i> / μmol L ⁻¹)
1	-1 (0.21)	-1 (2.4)	-1 (0.24)
2	-1 (0.21)	-1 (2.4)	+1 (0.81)
3	-1 (0.21)	+1 (8.1)	-1 (0.24)
4	-1 (0.21)	+1 (8.1)	+1 (0.81)
5	+1 (0.80)	-1 (2.4)	-1 (0.24)
6	+1 (0.80)	-1 (2.4)	+1 (0.81)
7	+1 (0.80)	+1 (8.1)	-1 (0.24)
8	+1 (0.80)	+1 (8.1)	+1 (0.81)
9	-1.68 (0.01)	0 (5.3)	0 (0.53)
10	+1.68 (1.0)	0 (5.3)	0 (0.53)
11	0 (0.51)	-1.68 (0.5)	0 (0.53)
12	0 (0.51)	+1.68 (10.0)	0 (0.53)
13	0 (0.51)	0 (5.3)	-1.68 (0.05)
14	0 (0.51)	0 (5.3)	+1.68 (1.0)
15	0 (0.51)	0 (5.3)	0 (0.53)
16	0 (0.51)	0 (5.3)	0 (0.53)

N-PLS modelling

All calculations were performed using MATLAB 7.0 (The MathWorks Inc.) for Windows and the N-way Toolbox for Matlab version 2.11 (obtained from R. Bro at <http://www.models.kvl.dk/source/>.³⁵). The software was validated using data sets available from <http://www.models.kvl.dk/source/nwaytoolbox/download.asp>. The files “claus.mat”, containing fluorescence excitation emission data from five samples of tryptophan, phenylalanine, and tyrosine; “brod.mat”, containing the assessments of eight judges over ten breads with respect to eleven attributes; and “sugar.mat”, with the fluorescence spectra of water-dissolved sugar, were run with the suite of programs used here and results verified against the published data.

Results and discussion

Osteryoung square wave voltammograms

The voltammograms prior to metal ion accumulation are typically featureless in the potential range between 0.40 and -0.45 V. Occasionally, small peaks due to metal ion contamination do arise in the voltammogram during the modification process or due to trace contamination in solutions. In this case, the currents of the blank voltammogram were subtracted from the voltammogram after accumulation. As a data preprocessing step, the difference voltammogram was also baseline subtracted prior to modelling. Illustrated in Fig. 3 are the baseline-subtracted voltammograms after accumulation of metal ions for 10 minutes at the four modified electrodes in calibration sample number 16 (the centre point of the central composite design which contains $0.51 \mu\text{M Cu}^{2+}$, $5.3 \mu\text{M Cd}^{2+}$ and $0.53 \mu\text{M Pb}^{2+}$). The OSW voltammograms obtained upon metal ion accumulation for all four modified electrodes displayed two peaks in the voltammogram. The more positive peak arising at ~ 0.1 V is due to the reduction of Cu^{2+} to Cu^0 . The more negative peak (in the region between -0.28 to -0.32 V) is due to both the reduction of Cd^{2+} to Cd^0 and Pb^{2+} to Pb^0 occurring at similar potentials. The reduction of Cu^{2+} , Cd^{2+} and Pb^{2+} results in the underpotential deposition of each metal onto the gold electrode surface.^{29, 30, 32} All four modified electrodes displayed different levels of metal uptake upon accumulation in Cu^{2+} , Cd^{2+} and Pb^{2+} . As expected, the greatest peak current density for copper ($5.0 \mu\text{A cm}^{-2}$) was obtained using the TA-Gly-Gly-His modified electrode (Fig. 3). This peptide is known to form a highly stable 4 N square planar complex.^{36, 37} The TA-Gly-Gly-His modified electrode was also able to bind Cd^{2+} and Pb^{2+} as illustrated by a smaller peak in the voltammogram at -0.315 V. For an electrode modified with TA only, smaller peak currents were obtained for Cu^{2+} , Cd^{2+} and Pb^{2+} compared to all other electrode types owing to the weaker binding capacity of the single carboxyl group.

These results using the four modified electrodes demonstrate the ability of peptides to bind metal ions with higher affinity than simple, unidentate ligands. The tuning of these amino acid sequences have led to sensors which display a broad range of selectivities for a range of metal ions. The resulting sets of voltammograms can then be used to calibrate Cu^{2+} , Cd^{2+} and Pb^{2+} using a suitable regression method such as N-PLS.

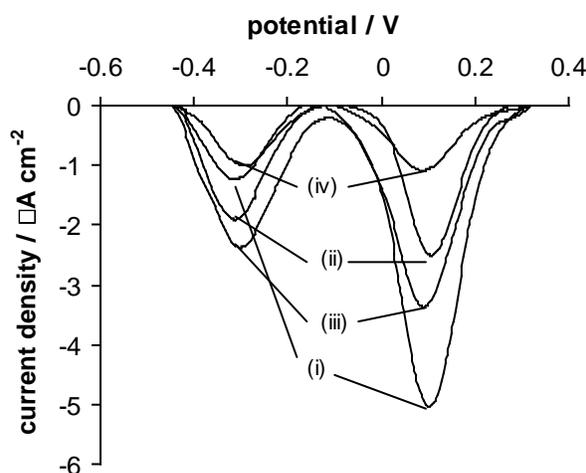


Figure 3: Baseline subtracted OSW voltammograms of the difference between the voltammograms obtained before and after metal ion accumulation in calibration sample number 16 (level (0, 0, 0) of Cu^{2+} , Cd^{2+} and Pb^{2+} in Table 1) in 50 mM ammonium acetate (pH 7.0) for 10 minutes at 25°C. The electrodes were modified with (i) TA-Gly-Gly-His, (ii) TA-glutathione, (iii) TA-angiotensin I and (iv) TA. OSW voltammograms were measured in 50 mM ammonium acetate (pH 7.0) and 50 mM NaCl at 25°C at a pulse amplitude of 0.025 V, a step of 0.004 V and frequency of 25 Hz.

N-PLS calibration and prediction

N-PLS was used to correlate the variations in the voltammogram with the variations in the concentrations of Cu^{2+} , Cd^{2+} and Pb^{2+} . Regression models were fitted for Cu^{2+} , Cd^{2+} and Pb^{2+} using 1 to 15 principal components (PCs) and the graphs in Fig. 4 illustrate the percentage of variation explained in the voltammogram currents and concentration with each PC. Fig. 4(a) shows an initial sharp rise in the percentage of variation explained in the sum of squares of the residuals of X (SS_X) and the sum of squares of the residuals of Y (SS_Y) for Cu^{2+} . This becomes almost constant after 2 PCs for both X and Y . For Cd^{2+} , most of the variation in the voltammogram currents could be fitted using 4 PCs and most of the variation in concentration could be fitted using 7 PCs (Fig. 4(b)). Similarly for Pb^{2+} , most of the variation in the voltammogram currents could be fitted using 2 PCs and most of the variation in concentration could be fitted using 5 PCs (Fig. 4(c)).

Prediction of the concentrations of Cu^{2+} , Cd^{2+} and Pb^{2+} for the calibration samples was made using the optimal number of PCs explaining the Y variance. These were 2, 7 and 5 PCs for Cu^{2+} , Cd^{2+} and Pb^{2+} , respectively. The standard error of

calibration ($\sqrt{\frac{SS_y}{N-p}}$, $N = 16$, $p =$ number of principal components) was 1.3×10^{-7} ,

7.0×10^{-7} and 8.8×10^{-8} M for Cu^{2+} , Cd^{2+} and Pb^{2+} , respectively. Fig. 5 shows the predicted concentrations *versus* the actual concentrations of the calibration solutions for

Cu^{2+} , Cd^{2+} and Pb^{2+} . The graph shows good correlations between the predicted and actual concentrations with the exception of samples at the extreme ends of the experimental design which show larger prediction errors. There is also some variation in prediction with the same analyte concentration from different samples due to the different extents of cross-interferences. To improve on the calibration of Cu^{2+} , Cd^{2+} and Pb^{2+} a more suitable range of concentrations may be necessary. The concentrations chosen for this study covered a wide dynamic range which can be problematic as calibration samples in the lower or upper concentration range (samples 8-14) may not fall in the linear range. Note that PLS is a linear method and thus requires more principal components to model the non-linear data over the entire range. The results also suggest that central composite design might not be the best for calibration as there are only single points at the extremities. A more appropriate calibration set for future work would have concentrations corresponding to a box design so that there are multiple points at the extremes.

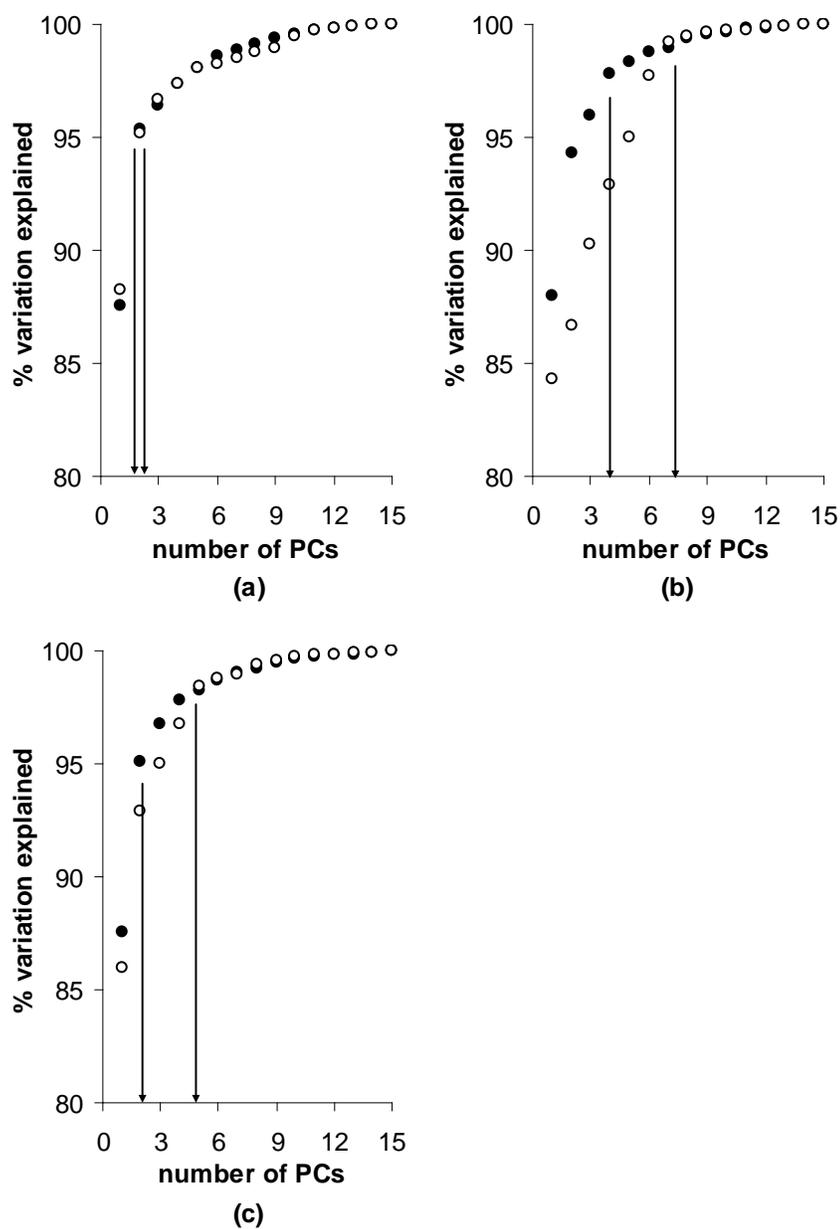


Figure 4: Percentage of variation explained in SS_x (●) and SS_y (○) after fitting a model to 1-15 PCs. X represents the voltammogram currents and Y represents the concentration of the analyte. The analytes are (a) Cu^{2+} , (b) Cd^{2+} and (c) Pb^{2+} . Arrows represent the optimal number of PCs.

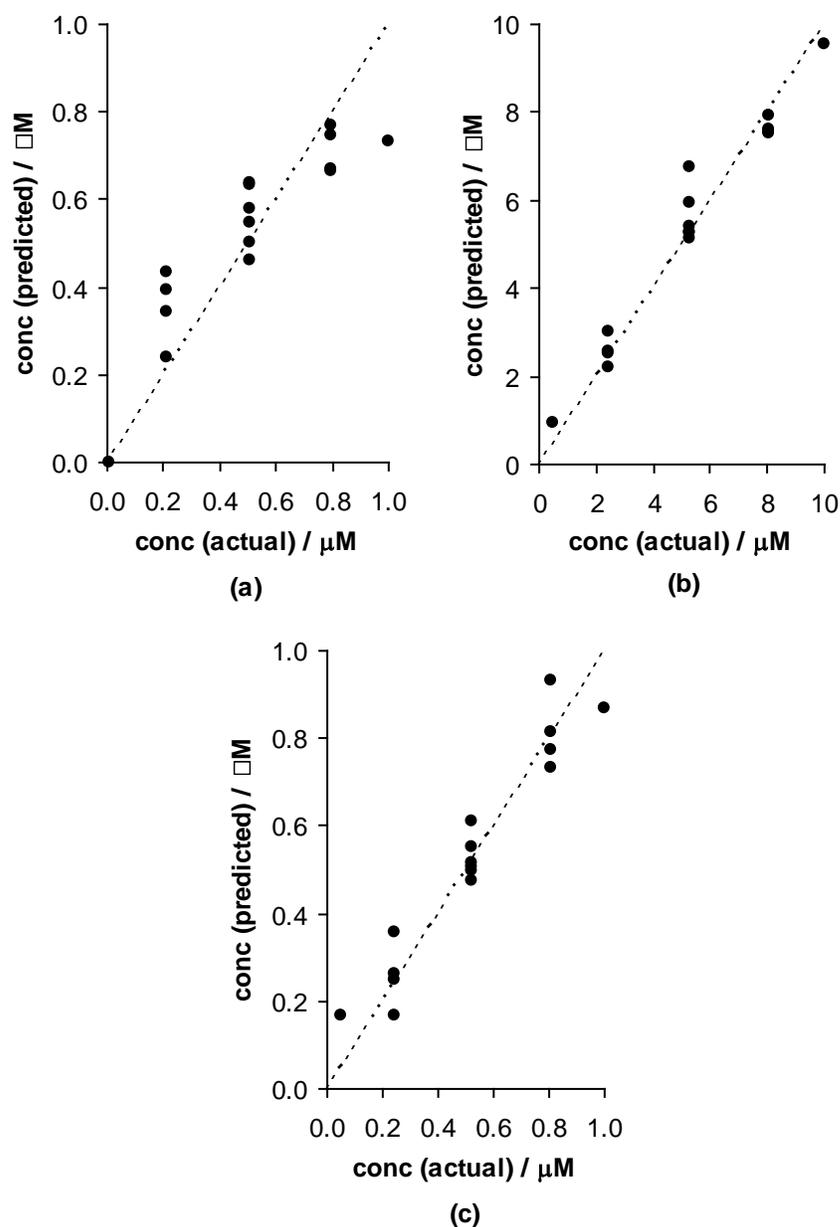


Figure 5: Predicted *versus* actual concentrations for samples 1-16 using N-PLS after fitting a model to 2, 7 and 5 PCs for (a) Cu^{2+} , (b) Cd^{2+} and (c) Pb^{2+} , respectively. The number of PCs used was determined from the Y variance explained in Fig. 4.

Prediction of three test samples was performed based on the calibrations for Cu^{2+} , Cd^{2+} and Pb^{2+} using 2, 7 and 5 PCs, respectively (Table 2). Without N-PLS modelling, the concentrations of Cu^{2+} , Cd^{2+} and Pb^{2+} in test samples would be very hard to predict from

the voltammograms. Not only do Pb^{2+} and Cd^{2+} voltammograms overlap, Pb^{2+} also reduces the copper peak in MPA-Gly-Gly-His when it competes for peptide.²⁹ For the prediction of Cd^{2+} in test sample 1, the signal may be masked by the high concentration of Pb^{2+} . The prediction error was largest at test samples where a particular analyte was at one of the extreme ends of the experimental design used for calibration, and reflect the nature of the central composite design. The deviations between actual and predicted concentrations are affected by the quality of the calibration set and the number of PCs chosen to explain the variance in the X and Y block.

Table 2 Prediction of Cu^{2+} , Cd^{2+} and Pb^{2+} in test samples using 2, 7 and 5 PCs, respectively.

	Analyte	Concentration (actual) / $\mu\text{mol L}^{-1}$	Concentration (predicted) / $\mu\text{mol L}^{-1}$
Test sample 1	Cu^{2+}	0.10	0.19
	Cd^{2+}	3.0	3.6
	Pb^{2+}	0.60	0.47
Test sample 2	Cu^{2+}	0.40	0.63
	Cd^{2+}	5.0	6.5
	Pb^{2+}	0.10	0.22
Test sample 3	Cu^{2+}	0.60	0.64
	Cd^{2+}	4.0	5.6
	Pb^{2+}	0.70	0.53

Comparison between N-PLS and PLS-1

Until the advent of multi-way methods, when data was obtained in more than two dimensions, it was customary to unfold the data into a single vector containing all information about a given object (sample). Although this loses the correlations between different dimensions, two-dimensional PLS has been shown to be a powerful calibration method³⁸. We calibrated the multielectrode system using PLS-1 and obtained comparable results. The standard errors of calibration for each ion were not significantly different between the two methods. This supports conclusions reached by Bro and others²³.

Comparison with reported methods

The method described here can be seen as complimentary to a number of stripping and data analytical approaches. Wang has shown that with under optimum conditions stripping analysis at a bare gold electrode can be effective for some combinations of metal ions (Cu^{2+} , Hg^{2+})⁴, and screen printed graphite, with and without mercury, can also detect Cu^{2+} , Cd^{2+} and Pb^{2+} ². An electrode modified with peptides has two advantages. First, the choice of peptide allows enhanced selectivity which also improves sensitivity and detection limit²⁷, and secondly, the peptide motif is a model of a bio-receptor, allowing estimation of metals that can be taken up by organisms. The N-way PLS calibration can be used with any system in which there are multianalytes and more than two data dimensions (here peptide x sweep potential), and it should provide a benefit, particularly as the systems become more complex, with overlapped signals.

Conclusions

The simultaneous determination of Cu^{2+} , Cd^{2+} and Pb^{2+} has been demonstrated using a four electrode sensor array. The results using TA-Gly-Gly-His, TA-glutathione, TA-angiotensin I and TA have shown how ligands can be tuned to display a broad range of selectivities for Cu^{2+} , Cd^{2+} and Pb^{2+} . N-PLS was used for calibration and prediction of a 16 sample set with good results for Cu^{2+} , Cd^{2+} and Pb^{2+} concentrations in the middle of the calibration and larger prediction errors for lower and higher concentrations. However, considering the extreme overlap of the voltammograms of lead and cadmium, the system has been successfully calibrated.

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Captions for schemes

Captions for figures

